



Full length article

Evaluation of the anti-oxidative and ROS scavenging properties of biomaterials coated with epigallocatechin gallate for tissue engineering



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ARTICLE INFO

Article history:

Received 2 November 2020

Revised 2 February 2021

Accepted 2 February 2021

Available online 6 February 2021

Keywords:

Tissue engineering

Oxidative stress

Reactive oxygen species

Antioxidant

Polyphenol coating

ABSTRACT

In tissue engineering, excessively generated reactive oxygen species (ROS) during biomaterial implantation or cell transplantation is a one of major causes of diminishing therapeutic effects. In this study, we prepared biomaterial surfaces coated with antioxidant epigallocatechin gallate (EGCG) and metal ions, and evaluated their anti-oxidative and ROS scavenging properties. We revealed that EGCG-coating on polycaprolactone (PCL) film surface increased hydrophilicity and anti-oxidative properties as a function of total phenol content (TPC) potentially due to the increase in phenolic -OH and π -electrons from structural maintenance and directly removed the hydrogen peroxide (H_2O_2) by resonance-stabilization. Furthermore, EGCG-coated PCL film increased attachment, spreading area, and viability of human adipose-derived stem cells (hADSCs) against H_2O_2 treatment while stimulated the cellular signaling to reduce apoptotic gene and enhance anti-oxidative enzyme expression. Further, we applied EGCG coating on the surface of poly-L-lactic acid (PLLA) fibers. Spheroids incorporating EGCG-coated PLLA fibers were able to maintain their shape and showed improved viability and anti-oxidative activities in response to H_2O_2 -induced oxidative stress than control spheroids. Therefore, metal-phenolic network (MPN) coating of EGCG is a suitable method to impart the anti-oxidative properties to biomaterials by evaluating the structural properties and biological effects.

Statement of Significance

This manuscript describes an antioxidant coating for biomaterials to control reactive oxygen species (ROS). Antioxidant epigallocatechin gallate (EGCG) was coated on the PCL film surface via metal-phenolic network (MPN). We revealed that the phenolic functional groups of EGCG are structurally maintained as confirmed by quantitative reducing power, radical scavenging assays. EGCG coating not only removed ROS directly, but also is involved in cell signaling to enhance the anti-apoptotic gene, anti-oxidative enzyme expression. Furthermore, human adipose-derived stem cells (hADSCs) spheroid produced by self-assembly with EGCG-coated poly-L-lactic acid (PLLA) fibers showed anti-oxidative properties from inside of spheroids. Thus, our evaluation of the anti-oxidative properties of EGCG coating can be applied to various tissue engineering applications.

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1. Introduction

Reactive oxygen species (ROS), including superoxide ($\bullet O_2^-$), hydrogen peroxide (H_2O_2), and hydroxyl radicals ($\bullet OH$), are generally produced during implantation of biomaterials or cells for tissue

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engineering therapy. Excessive ROS cause apoptosis and activation of inflammatory signaling cascades, leading to incomplete recovery of damaged tissue [1,2]. For example, the high level of ROS surrounding dental implants results in uncontrolled inflammation with bone loss, and elevation of ROS during autologous or allogenic hematopoietic stem cell transplantation (HSCT) can result in serious toxic problems [3–5]. It has been reported that pro-oxidant methemoglobin (Fe^{3+}) released from lysed blood cells and H_2O_2 derived from the lysis of epithelial or immune cells during surgical incision of implantation process are associated with ROS-mediated failure of tissue engineering approaches [6,7]. In addition, serum proteins adsorbed to the surfaces of biomaterials are known to bind to Fc domains of transmembrane receptors of inflammatory cells, stimulating the synthesis of ROS by these cells [8,9]. Therefore, it is important to control ROS for successful regenerative outcomes in tissue engineering.

Various strategies based on the use of antioxidants and anti-oxidative enzymes have been investigated to control ROS. For example, delivery of the soluble antioxidant N-acetylcysteine (NAC) with mesenchymal stem cell (MSC) aggregates into ischemic tissue successfully removed 80% of H_2O_2 with enhancement of cell adhesion and survival of the engrafted cells [10]. Furthermore, delivery of the soluble antioxidant edaravone with human umbilical cord mesenchymal stem cells (hUMSCs) in an acute liver failure model increased the viability of transplanted hUMSCs by 1.5-fold and reduced the area of hepatic necrosis by approximately 60% [11]. However, co-injected soluble antioxidants or anti-oxidative enzymes were rapidly cleared (8–90% within 24 hr), and therefore, ineffective for continuous control of ROS generated by damaged tissue [12,13]. As an alternative, biomaterials have been mixed or surface-coated with antioxidant or anti-oxidative enzymes to achieve sustained anti-oxidative activity. For example, polycaprolactone (PCL)/polyethylene glycol (PEG) electrospun nanofibers mixed with antioxidant chrysin significantly reduced chrysin release with 1,000-fold greater oxygen radical decomposition capacity than control nanofibers, while hyaluronic acid hydrogels conjugated with anti-oxidative dopamine showed 40% and 65% improved hydroxyl radical and DPPH scavenging activity over non-conjugated hydrogels, respectively, with increased SOD and GPX enzyme activity [14,15]. However, mixing of ROS-controlling agents with biomaterials may alter the characteristics of biomaterials such as their degradation rate, swelling ratio, and mechanical properties, while the surface conjugating strategy may require complex chemical processes and could decrease the biological activities of antioxidants or anti-oxidative enzymes [14–16].

Polydopamine and polyphenol coating of biomaterials have been actively studied. Polydopamine forms a material-independent coating on the biomaterial surface at a basic pH by self-oxidation of catechol groups in dopamine [17]. Similarly, polyphenols such as tannic acid (TA), catechin (CA), and epigallocatechin gallate (EGCG) can readily form a metal-phenolic network (MPN) with cationic metal ions due to hydrogen bonding, π - π stacking, and cation- π interactions by incubation of the sample in a buffer solution containing metal ions such as Na^+ , Fe^{3+} [18]. In particular, an MPN can form on the surfaces of various types of materials, facilitating simple and effective coating and conferring that material with the biological properties of polyphenols including their anti-inflammatory, anti-cancer, and anti-oxidative properties [19]. For example, gold nanoparticles coated with polyphenols in the presence of $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ showed 46%, 51%, and 55% ABTS radical scavenging activity when coated with EGCG, resveratrol (RSV), and fisetin (FS), respectively [20]. Furthermore, tissue culture polystyrene (TCP) coated with TA modified with Na^+ coated silica nanoparticles reduced the intracellular ROS level significantly more than uncoated TCP [21]. In addition, MPN coatings have been reported to have anti-cancer activity (coating of RSV with Au^{3+})

and to enhance endothelialization (coating of TA with Sr^{2+}) [22–24]. Despite the focus on using polyphenols as surface modifiers of biomaterials, detailed information on the effects of coating concentration and structural feature of polyphenols on anti-oxidative activity and cellular signaling have not yet been reported.

Previously, our group reported the EGCG coating on Ti surface in the presence of Mg^{2+} , and investigated their effect on osteogenic differentiation of human adipose-derived stem cells (hADSCs) and suppression of osteoclast functions [25]. Compared to this previous work, the current study is focused on the evaluation of the anti-oxidative properties of EGCG-coated biomaterial surfaces and their effects on cells, based on the structural and chemical properties of EGCG through 2D and 3D culture conditions for hADSCs. Given that, we prepared PCL films and poly-L-lactic acid (PLLA) fibers coated with EGCG as model substrates, and then, analyzed the effect of coating in the presence of cations (Na^+ or Mg^{2+}) on the total phenol content (TPC) deposited on the surfaces of the substrates, surface properties, and anti-oxidative properties. Second, we cultured hADSCs on EGCG-coated surfaces and investigated the effect of the EGCG coating on the survival of hADSCs under a ROS environment induced by H_2O_2 treatment. In addition, PLLA fibers coated with EGCG were combined with hADSCs to prepare 3D spheroids and we examined whether the EGCG-coated fibers were able to protect stem cells within the spheroids from oxidative stress.

2. Materials and methods

Polycaprolactone (PCL) ($M_w=80,000$) and poly-L-lactic acid (PLLA, 1479) were obtained from Sigma-Aldrich (St. Louis, MO, USA) and Samyang (Jeollabuk-do, Korea), respectively. 2,2,2-trifluoroethanol (TFE) and ethylenediamine (EDM) were purchased from Sigma-Aldrich (St. Louis, MO, USA). Dichloromethane (DCM) and isopropyl alcohol (IPA) were purchased from Junsei Chemical Co., Ltd (Chuo-ku, Tokyo, Japan) and EDM Millipore (Darmstadt, Germany), respectively. BIS-TRIS was obtained from Sigma-Aldrich (St. Louis, MO, USA), and EGCG was purchased from TCI America. Magnesium chloride hexahydrate ($\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$) and sodium chloride (NaCl) were obtained from Junsei Chemical Co., Ltd. Distilled water (D.W.) was acquired from an Elix Advantage System (Millipore, MA, USA). Phosphate buffered saline (PBS), trypsin/EDTA, fetal bovine serum (FBS), and penicillin-streptomycin (PS) were purchased from Wisent (St. Bruno, QC, Canada). MesenPro RS medium and L-glutamine were obtained from Gibco BRL (Rockville, MD, USA). Human adipose-derived stem cells (hADSCs) were purchased from Invitrogen (Carlsbad, CA, USA). 3% hydrogen peroxide (H_2O_2) was purchased from Sigma-Aldrich (St. Louis, MO, USA). LIVE/DEAD assay kit and Alexa FluorTM 488 Phalloidin were purchased from Invitrogen (Carlsbad, CA, USA). Mounting medium containing 4',6-diamidino-2-phenylindole was purchased from Vectashield® (Burlingame, CA, USA). The RNeasy Mini Kit was obtained from Qiagen (Valencia, CA, USA). Maxime RT Premix was purchased from Intron (Seoul, Korea), and SYBR Premix Ex Taq was acquired from TAKARA (Otsu, Shiga, Japan).

2.1. Preparation and characterization of EGCG-coated PCL films

To prepare PCL films, PCL granules (4 g) were dissolved in 50 ml of dichloromethane (DCM) and 2,2,2-trifluoroethanol (TFE) (8:2, v/v) for 24 hr, and this solution was then poured into a 100 mm glass dish and thoroughly dried in a 60°C dry oven for 6 hr. Circles were punched from the dried PCL film (diameter, 1.91 cm²). 1 mM EGCG solution was prepared in BIS-TRIS buffer (100 mM of BIS-TRIS in D.W. at pH 7). We added either 300 mM of NaCl or MgCl_2 to the buffer solution for MPN formation. PCL film was placed on a 24-well plate and incubated with 1 ml of EGCG solution for 24 hr

at 37°C. The total phenol content (TPC) on the PCL films was determined using the Folin-Ciocalteu method. Briefly, films were incubated with 300 μ L Folin-Ciocalteu solution for 10 min, and then 700 μ L of sodium carbonate solution (Na_2CO_3 , 2% w/v) was added. After a 2-hr incubation at RT, absorbance was measured at 780 nm. The TPC on the PCL film was calculated using an EGCG standard. The same concentration (300 mM) of buffer solutions containing different monovalent (NaCl , KCl) and divalent (MgCl_2 , CaCl_2 , and SrCl_2) metal ions were used for the EGCG coating on PCL film surface with same method described above. The relative TPC on PCL film was calculated by using the same Folin-Ciocalteu method and normalization with the values from the use of NaCl and MgCl_2 , respectively. The EGCG-coated PCL films were incubated in 1 ml of D.W. for 1, 2, 4, and 7 days to measure the remaining TPC on the surface. The relative TPC of Surface functional groups on the PCL film were analyzed by attenuated total reflection-Fourier transform infrared spectroscopy (ATR-FTIR, Nicolet 6700, Thermo Scientific, Waltham, MA, USA) and the surface atomic composition of PCL film was analyzed by X-ray photoelectron spectrometry (XPS) (ThetaProbe Base System, Thermo Fisher Scientific) at the Hanyang Center for Research Facilities (Seoul) using analysis software (Thermo VG Scientific, MA, USA). Surface morphological changes were observed using scanning electron microscopy (SEM) (Hitachi S-4800, Hitachi, Ltd, Tokyo, Japan). Water contact angle was measured by capturing images (Phoenix 300, Surface Electro Optics Co., Suwon, Korea) of a 1 μ L droplet of D.W. on the surface of the PCL films and analyzing them with Image-Pro plus software (MediaCybernetics, Silver Spring, MD, USA).

Anti-oxidative properties of EGCG-coated PCL films were analyzed using a ferric reducing antioxidant power assay and ABTS radical scavenging assay. For the ferric reducing antioxidant power assay, 1 mg/ml of 1,10-phenanthroline and 1 mM of FeCl_3 (Sigma-Aldrich, St. Louis, MO, USA) were dissolved in D.W. to obtain an Fe(III) solution. Prewetted EGCG-coated PCL films were mixed with 1 ml of Fe(III) solution, and the absorbance of the solution was measured at 510 nm and used to quantify the conversion ratio of Fe(III) to Fe(II) (%) calculated using an ascorbic acid standard. ABTS radicals were generated in a solution containing 7.0 mM 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt and 2.4 mM potassium persulfate ($\text{K}_2\text{S}_2\text{O}_8$) (Sigma-Aldrich, St. Louis, MO, USA) in D.W. for 24 hr. The ABTS solution was diluted with PBS until an absorbance value of 0.7 at 732 nm was reached and then mixed with 50 μ L of ascorbic acid standard or EGCG-coated PCL film prewetted with 50 μ L D.W. After 10 min of reaction, the percentage ABTS radicals scavenged was calculated by measuring the absorbance of the solution at 732 nm.

2.2. Adhesion of hADSCs and ROS-scavenging activity of EGCG-coated PCL films

hADSCs were maintained under standard culture conditions with MesenPro RS medium (with 1% L-glutamine and 1% PS) and cells from passage number 4 were used in all experiments. To determine the effect of EGCG on the viability of hADSCs, hADSCs were seeded in a tissue culture plate at 0.5×10^4 cells/ cm^2 and cultured for 24 hr. Then, the media was replaced with fresh medium containing EGCG (0, 12.5, 25, 50 μ M) and the viability of hADSCs was measured by MTT assay after a 24-hr incubation. hADSCs were seeded at 0.5×10^4 cells/ cm^2 on the surface of PCL film and EGCG-coated PCL film and cultured for 24 hr. hADSCs cultured on PCL film were fixed with 4% paraformaldehyde, and then incubated with 1:200 Alexa FluorTM 488 Phalloidin for 1 hr at 37°C for F-actin staining. hADSCs stained for F-actin were preserved using mounting medium with DAPI. The number of attached cells was calculated by counting the number of nuclei per area (mm^2), and spread area was calculated by measuring the existing area ($\times 10^3$

μm^2) of actin filaments per individual cells using Image-Pro plus software.

We first investigated the effect of the concentration of exogenous H_2O_2 on the viability of hADSCs; hADSCs were cultured on a tissue culture plate at 0.5×10^4 cells/ cm^2 for 24 hr and the medium was replenished by fresh medium containing 0, 100, 200, or 400 μ M H_2O_2 . Cell viability was then measured by MTT assay after 4, 8, and 24 hr. To investigate the effect of EGCG on ROS scavenging, hADSCs were similarly cultured on a tissue culture plate at 0.5×10^4 cells/ cm^2 , and the culture media was then replaced by one containing H_2O_2 (100 μ M) and 0, 12.5, 25, or 50 μ M of EGCG. After 24 hr of culture, cell viability was measured by MTT assay. We then cultured hADSCs on the surfaces of PCL film and EGCG-coated PCL film for 24 hr and exposed the hADSCs on the film to 200 μ M of H_2O_2 -containing culture media for 12 hr. The number of cells before and after H_2O_2 treatment were counted and morphological changes were observed after F-actin staining.

2.3. Anti-apoptotic and anti-oxidative activities of EGCG-coated PCL film against H_2O_2

Apoptotic DNA fragmentation of hADSCs cultured on PCL and EGCG-coated PCL films in response to H_2O_2 treatment was analyzed by TUNEL assay (ApopTag[®] fluorescein in situ apoptosis detection kit, Sigma-Aldrich, St. Louis, MO, USA). hADSCs at 0.5×10^4 cells/ cm^2 were cultured on the surfaces of PCL and EGCG-coated PCL films for 24 hr, and the films were then exposed to 200 μ M of H_2O_2 for 12 hr. H_2O_2 -exposed cells on PCL films were fixed with 1% paraformaldehyde for 10 min and then treated with ethanol:acetic acid (2:1, v:v) for 5 min. After two washes with PBS, equilibration buffer was added for 10 sec. Then, the cells were incubated with terminal deoxynucleotidyl transferase (TdT) for 1 hr in a 37°C incubator. Immediately after TdT treatment, stop/wash buffer was added for 10 min at RT, followed by addition of anti-digoxigenin conjugate (fluorescein) for 30 min. Stained cells were preserved in mounting medium with DAPI. Percentage TUNEL-positive nuclei was calculated by counting the number of fluorescein-stained nuclei (positive) and normalizing this to the total number of nuclei.

To examine the ability of EGCG-coated film to scavenge H_2O_2 , PCL and EGCG-coated PCL films were incubated in D.W. containing 100 μ M H_2O_2 for 2 hr and the remaining H_2O_2 was measured using an H_2O_2 detection kit (Fluorimetric Hydrogen Peroxide Assay Kit, Sigma-Aldrich, St. Louis, MO, USA). Cells were also cultured as described above and then treated with H_2O_2 for 12 hr, followed by incubation with 10 μ M of 2',7'-dichlorofluorescein diacetate (DCFH-DA) (Sigma-Aldrich, St. Louis, MO, USA) for 30 min. Intracellular ROS level was assessed by fluorescence microscopy, and the relative mean fluorescence intensity (MFI) value was quantified by calculating fluorescence intensity within areas of individual cells using Image-Pro plus software and normalized to that measured in the PCL group. We also analyzed the expression of apoptotic and oxidative genes by real-time reverse transcription polymerase chain reaction (RT-PCR). Total RNA of hADSCs cultured on EGCG-coated PCL film was collected using TRizol reagent (Life Technologies, Carlsbad, CA, USA), and cDNA was synthesized from collected total RNA using the Maxime RT PreMix Kit (Intron Biotechnology, Gyeonggi-do, Korea). Then, the cDNA was mixed with SYBR Green PCR Mastermix (TAKARA, Otsu, Shiga, Japan), and PCR was performed in a StepOnePlus thermocycler (Life Technology, Carlsbad, CA, USA). The sequences of the primers used for RT-PCR were as follows: GAPDH (Fw: 5'-GTC AGT GGT GGA CCT GAG CT-3', Rv: 5'-TGC TGT AGC CAA ATT CGT TG-3'), BAX (Fw: 5'-TTT GCT TCA GGG TTT CAT CC-3', Rv: 5'-CAG TTT AAG TTG CCG TCA GA-3'), BCL2 (Fw: 5'-GAG GAT TGT GGC CTT TTG TG-3', Rv: 5'-ACA GTT CCA CAA AGG CAT CC-3'), BCL2L1 (Fw: 5'-CTG AAT CGG AGA TGG AGA CC-3', Rv:

5'-TGG GAT GTC AGG TCA CTG AA-3'), Catalase (Fw: 5'-GCC TGG GAC CCA ATT ATC TT-3', Rv: 5'-GAA TCT CCG CAC TTC AG-3'), FOXO3 (Fw: 5'-GCA AGC ACA GAG TTG GAT GA-3', Rv: 5'-CAG GTC GTC CAT GAG GTT TT-3'), GPX-1 (Fw: 5'-CTC TTC GAG AAG TGC GAG GT-3', Rv: 5'-TCG ATG TCA ATG GTC TGG AA-3').

2.4. Preparation and characterization of EGCG-coated PLLA fibers

To prepare PLLA fibers, an electrospun nanofiber sheet was prepared by ejecting 10 ml of 3% of PLLA solution dissolved in a mixture of DCM and TFE (8:2, v/v) using a syringe pump (KDS200, KD Scientific, New Hope, PA, USA) through a 23-gauge needle under 16 kV at a rate of 2 ml/h. Scattered fibers were collected on aluminum foil covered a rotating mandrel (SPG, Incheon, Korea), and the sheet was dried overnight. The nanofiber sheet was chopped into small pieces and dispersed in ethylenediamine (EDM) solution (10%, v/v in IPA) for aminolysis. After vigorous shaking at 90 rpm for 30 min at 37°C, the aminolyzed PLLA fibers were centrifuged at 4,000 rpm for 10 min and washed in 70% ethanol. 50–100 µm sized PLLA fibers were collected using a sieve grid and lyophilized after 3 D.W. washes. To coat PLLA fibers (PF) with EGCG, 1 mg/ml of prepared PF (50–100 µm) was dispersed in EGCG solution (100 mM of BIS-TRIS with 300 mM of MgCl₂ dissolved in D.W. at pH 7) and incubated for 24 hr at 37°C on a rotator. The TPC on the PLLA fibers was measured by the Folin-Ciocalteu method as described in Section 2.1.

The anti-oxidative activity of EGCG-coated PLLA fibers (E-PF) was measured by a ferric reducing antioxidant power assay, ABTS radical scavenging assay, and H₂O₂ scavenging assay. PF and E-PF were dispersed in D.W. (1 mg/ml) and serially diluted. 500 µl of the PF sample, E-PF sample, and ascorbic acid standard were reacted with Fe(III) solution for 30 min and the absorbance was measured at 510 nm. Similarly, 50 µl of the PF sample, E-PF sample, and ascorbic acid standard were reacted with ABTS solution and the absorbance was measured at 732 nm. To analyze H₂O₂ scavenging activity, PF and E-PF were dispersed in 100 µM H₂O₂ solution (0 to 1 mg/ml), dissolved in D.W., and incubated for 2 hr at 37°C. The remaining concentration of H₂O₂ in the solution was measured using an H₂O₂ detection kit. To evaluate the biocompatibility of PF and E-PF and hADSCs, hADSCs were cultured on a 24-well plate at 1×10^4 cells/cm² for 24 hr. Then, the media was changed to one containing 5 µg of PF or E-PF. After 24 hr of culture, the viability of hADSCs was measured by MTT assay. The intracellular ROS level of hADSCs was measured by incubating the cells with 10 µM DCFH-DA for 30 min after the same culture process as described above. Fluorescence intensity of individual cells was calculated by Image-Pro plus software and normalized to that of the control (-) group.

2.5. hADSC spheroid formation with fibers and anti-oxidative effect of EGCG coated fibers on H₂O₂-treated hADSCs

To fabricate hADSC spheroids (cells only), 40,000 hADSCs in 100 µl of culture media were placed in a 0.2 ml tube and centrifuged at 1,200 rpm for 3 min. Cells in the tube gradually formed a spheroid during incubation for 24 hr at 37°C, and the spheroid was then transferred to an ultra-low attachment plate (Costar® ultra-low attachment multiple 96-well plate, Corning Incorporated, New York, USA). PF-incorporating spheroids were prepared as described above except that 10 µg of PF (1 µg/µl) was added the 100 of culture medium containing 40,000 cells. Cell-only spheroids and PF-incorporating spheroids were cultured in media containing 400 µM H₂O₂ for 12 hr. Changes in the morphology of spheroids were monitored by phase contrast microscopy and the number of deformed spheroids was quantified by image analysis and reported as a percentage. Using the same cell culture process described

above, the distribution of live and dead cells in spheroids was determined by confocal microscopy (TE2000 and Eclipse C1, Nikon, Tokyo, Japan) after 30 min of incubation in LIVE/DEAD staining solution (Invitrogen, Carlsbad, CA, USA).

For histological analysis, H₂O₂-exposed spheroids were fixed in 4% paraformaldehyde. Fixed individual spheroids were then immersed in optimal cutting temperature compound (OCT) solution and frozen in a deep freezer (-80°C) for more than 2 hr to produce an OCT block. The frozen OCT block containing spheroids was cut into 10 µm sections using a cryostat microtome and sections were collected on slide glass. To observe the internal morphology of spheroids, H&E staining of cross-sectioned spheroids was performed. First, cross-sectioned samples were incubated twice in xylene for 10 min. After a hydration step (rinsing with 100% to 70% ethanol followed by running water for 10 min), samples were incubated in hematoxylin for 2 min and eosin for 8 min. Following dehydration, samples were preserved in mounting medium for observation. The DNA integrity of hADSCs within spheroids was determined by TUNEL assay. First, cross-sectioned samples of spheroids from the OCT block were incubated twice in xylene for 10 min. Following hydration, samples were treated with ethanol:acetic acid (2:1, v:v) for 5 min. After two washes with PBS, samples were incubated in equilibration buffer for 10 sec. Then, samples were incubated with terminal deoxynucleotidyl transferase (TdT) for 1 hr at 37°C. Immediately after TdT treatment, stop/wash buffer was added for 10 min at RT, followed by anti-digoxigenin conjugate (fluorescein) for 30 min. Stained samples were preserved in mounting medium with DAPI. TUNEL-positive nuclei (%) were quantified by counting the number of fluorescein-stained nuclei (positive) normalized to the total number of nuclei. In addition, spheroids exposed to 400 µM H₂O₂ for 12 hr were incubated in 10 µM DCFH-DA for 30 min, and then intracellular ROS levels were quantified by fluorescence microscopy. Relative MFI was quantified by calculating the fluorescence intensity of individual spheroids and normalizing this to that of cell-only spheroids. For apoptotic and oxidative gene expression analyses, spheroids exposed to H₂O₂ were immersed in TRIzol reagent, and were then broken apart by pipetting several times to extract RNA. Subsequent steps for RT-PCR were the same as those described in Section 2.3.

2.6. Statistics and analyses

Quantitative data are presented as means ± standard deviations. One-way analysis of variance (ANOVA) using Tukey's honestly significant difference test and two-way ANOVA (for two variables) using the Bonferroni post-test were performed using GraphPad Prism 5 software (La Jolla, CA, USA). A *p* value < 0.05 was considered to denote statistical significance.

3. Results and discussion

3.1. Characterization of EGCG-coated PCL films

The overall scheme of EGCG coating on a biomaterial surface through MPN formation and the potential effects on 2D, 3D cultured cell underlying its anti-oxidative activity are illustrated in Fig. 1. According to the EGCG coating, the white color PCL surface turned slightly brown in Na-E and Mg-E, and small aggregates were observed in Mg-E (Supplementary Fig. S1). The TPC on the PCL film increased significantly in the presence of cations, with Mg²⁺ increasing TPC on the PCL film to a greater extent than Na⁺ (17 ± 3 µg/cm² vs 8 ± 1 µg/cm², respectively) (Fig. 2a). When various types of metal ions were used for EGCG coating, TPC from divalent was significantly greater than that from monovalent ions while no significant difference was found in different ions of each group (Supplementary Fig. S2). The TPC on PCL film

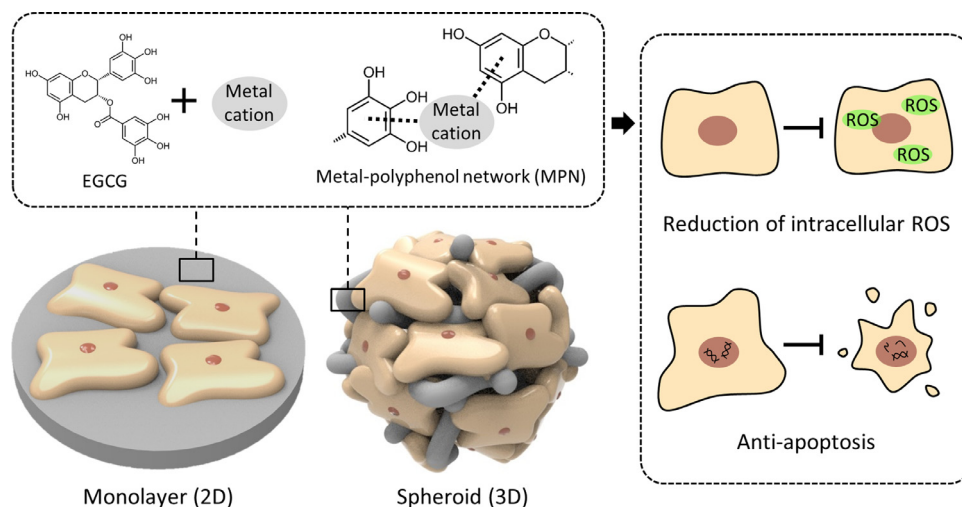


Fig. 1. Schematic diagram of the anti-oxidative properties of EGCG coating to reduce intracellular ROS and anti-apoptosis on 2D, 3D cultured cells.

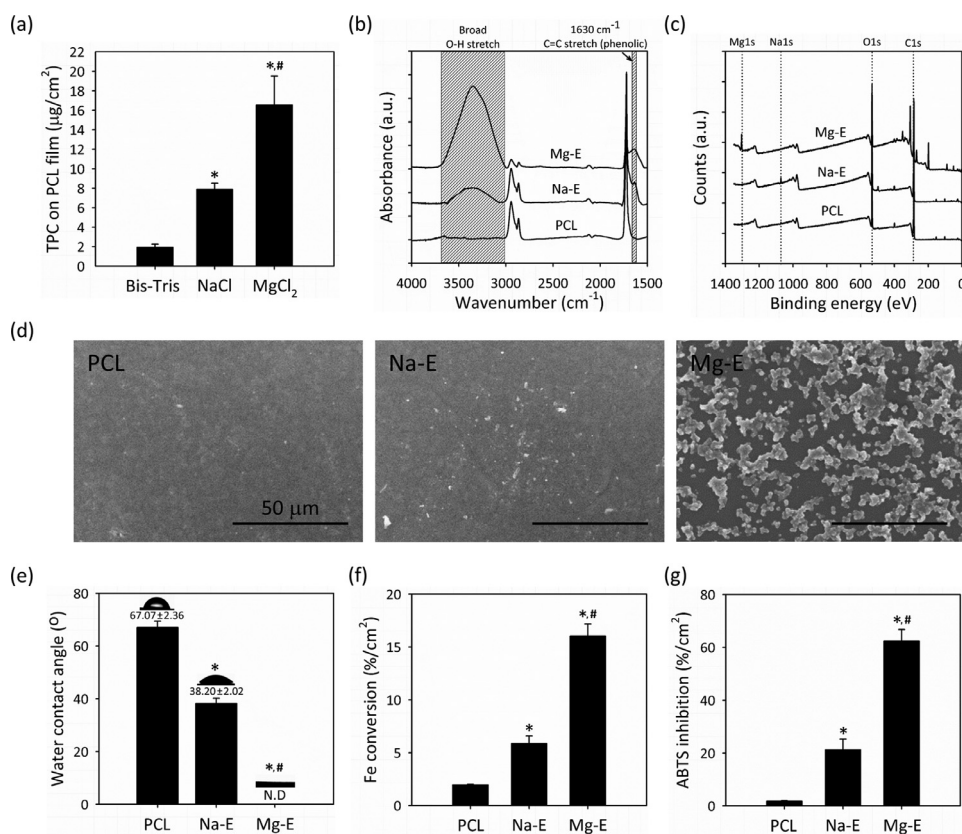


Fig. 2. Characterization of EGCG-coated PCL film. (a) The TPC on the PCL film surface using NaCl- or MgCl₂-containing buffer. Spectra of PCL film, Na-E, and Mg-E obtained by (b) ATR-FTIR analysis and (c) XPS analysis. (d) SEM images of the surfaces of PCL film, Na-E, and Mg-E. (e) Water contact angles of PCL film, Na-E, and Mg-E. The anti-oxidative activity of EGCG-coated PCL film as measured by (f) ferric reducing antioxidant power assay and (g) ABTS radical scavenging assay (*, # $p < 0.05$).

surface remained 80% and 60% in Na-E ($6 \pm 1 \mu\text{g}/\text{cm}^2$) and Mg-E ($10 \pm 1 \mu\text{g}/\text{cm}^2$) for 7 days (Supplementary Fig. S3). ATR-FTIR results demonstrated that a broad O-H stretch at $3000\text{--}3700 \text{ cm}^{-2}$ and phenolic C=C stretch at 1630 cm^{-1} were present after coating with EGCG (Fig. 2b), indicating successful coating with EGCG. These results are consistent with a previous report [26]. In addition, specific peaks of Na1s and Mg1s were detected by XPS analysis on the surface of Na-E and Mg-E, respectively (Fig. 2c). In the XPS C1s high-resolution spectra, peak intensity of C-O bonding (286.4 eV) was elevated in Na-E and more in Mg-E than PCL (Supplementary Fig. S4). SEM results showed that bare PCL had a

smooth surface while agglomerated particles were found on the Mg-E surface (Fig. 2d). Following EGCG coating, bare PCL film ($67 \pm 2^\circ$) became hydrophilic and complete wetting was observed on the PCL surface coated with the greater TPC (Fig. 2e). As shown in Fig. 2f and g, the conversion ratio of Fe ($\%/ \text{cm}^2$) was significantly elevated after EGCG coating as compared to bare PCL ($2 \pm 0\%/ \text{cm}^2$), and Mg-E ($16 \pm 1\%/ \text{cm}^2$) had a significantly greater Fe conversion ratio than Na-E ($6 \pm 1\%/ \text{cm}^2$). Similarly, Mg-E inhibited ABTS radicals significantly more than bare PCL and Na-E (Fig. 2g).

The divalent cation Mg^{2+} led to a greater level of MPN-based coating than the monovalent cation Na^+ . MPN is the supramolec-

ular network structure, in which metal ions are coordinated to phenolic ligands with π -cation interactions playing a crucial role [19,27]. In a previous study, no catechin (CA) coating was formed on a PCL nanofiber surface in the absence of sodium ion (Na^+) [28]. Similar to our findings, more pyrogallol (PG) coating was achieved on the titanium surface with magnesium ion (Mg^{2+}) than Na^+ [29]. In the XPS C1s high-resolution results, the gradual improvement in C-O bonding in Na-E and Mg-E than in PCL can be estimated by the improvement of phenolic C-O contents due to the increase in TPC of the surface. The structure of MPN with non-covalent interactions between metal ions and phenolic molecules has been indirectly estimated by stoichiometry and XPS analysis [19,30]. For example, mono/bis/tris-complex was estimated through the relative ratio of Fe^{3+} and TA, and the peak shift to higher binding energy through XPS analysis would have caused electron transfer from TA to Fe ions. In our results, the increased phenolic C-O content indirectly shows that Mg^{2+} can form more MPN than Na^+ . SEM analysis consistently supported more coating with Mg^{2+} than Na^+ . The formation of small aggregates was reported during polyphenol coating when coating exceeded a certain level [29]. The surface hydrophilicity increased with increase in TPC, indicating that the -OH groups of catechin and gallol in EGCG after coating may have contributed to the formation of hydrogen bonds with H_2O on the surface and the increased hydrophilicity. Consistently, the Mg-E surface exhibited 2–3-fold greater Fe conversion/ABTS scavenging activity than Na-E surface, suggesting that phenolic contents on the surface appear to be responsible for antioxidant activity to reduce Fe ($\text{Fe}^{3+} \rightarrow \text{Fe}^{2+}$) and to scavenge radicals [31]. It has been reported that the hydroxyl groups in phenolic ring structure of EGCG can scavenge radicals and chelate metal ion during self-oxidation, and the electron transfer in phenolic π -electrons is dependent on the amount of EGCG coated on the surface of the substrate [32].

3.2. Adhesion of hADSCs and ROS scavenging activity of EGCG-coated PCL films

To confirm the viability of hADSCs after EGCG treatment, cells were observed after LIVE/DEAD staining and cellular metabolic activity was quantified by MTT assay (Fig. 3a and b). Cell morphology and viability were not significantly affected by EGCG treatment. EGCG has been reported to be cytotoxic to mesenchymal cells only at concentration higher than 50 μM [33]. Consistent with this previous study, EGCG at concentrations lower than 50 μM was not cytotoxic to hADSCs. The number of attached cells on films cultured for 24 hr was significantly greater on the EGCG-coated surfaces (Na-E, $177 \pm 21/\text{mm}^2$ and Mg-E, $128 \pm 7/\text{mm}^2$) than on the PCL film ($87 \pm 20/\text{mm}^2$) (Fig. 3c). In addition, hADSCs became narrower and thinner (spreading area $5.2 \pm 1.6 \times 10^3 \mu\text{m}^2$ per cell) on the PCL film, while they had a more broad polygonal shape on Na-E and Mg-E films (spreading area $9.0 \pm 3.5 \times 10^3 \mu\text{m}^2$ and $7.3 \pm 1.5 \times 10^3 \mu\text{m}^2$ per cell, respectively) (Fig. 3d and e). According to previous reports, surface characteristics such as hydrophilicity and roughness are altered by polyphenol coating, and cell attachment and spreading area change accordingly. For example, the Fe^{3+} /TA coated surface of PCL nanofiber mesh became hydrophilic and small aggregates formed on the surface, similar to our findings [34]. Furthermore, human umbilical vein endothelial cells (HUVECs) cultured on PCL nanofiber mesh showed greater adhesion and spreading than HUVECs cultured on uncoated PCL nanofibers [34]. The reduced number of adherent cells on the Mg-E surface may be due to the roughness of the Mg-E surface. It has been reported that the adhesion force of cells decreases as surface roughness increases above a certain level (adhesion inhibition) [35].

Viability of hADSCs after H_2O_2 treatment decreased significantly as the concentration of H_2O_2 increased and exposure time increased (Fig. 4a). For example, after 24 hr, the viability of hADSCs decreased from $83 \pm 2\%$ at 200 μM H_2O_2 to $11 \pm 0\%$ at 400 μM H_2O_2 compared to the control. However, when cells were treated with both H_2O_2 and EGCG (0, 12.5, 25, 50 μM), the viability of hADSCs after 24 hr improved significantly (by ~ 10 –20%) as compared to that without EGCG ($74 \pm 5\%$) (Fig. 4b). Changes in the morphology of hADSCs cultured on PCL film and EGCG-coated PCL film were observed after 12 hr of exposure to 200 μM H_2O_2 . Cells cultured on PCL film were shrunken and had a low spread area ($3 \pm 2 \times 10^3 \mu\text{m}^2$ per cell) while cells cultured on Na-E and Mg-E exhibited an increased spread area relative to cells cultured on PCL film ($12 \pm 3 \times 10^3 \mu\text{m}^2$ and $11 \pm 3 \times 10^3 \mu\text{m}^2$ per cell, respectively) (Fig. 4c and d).

EGCG treatment protected hADSCs against H_2O_2 mediated-ROS damage, consistent with a previous study that reported that 75% of hMSCs treated with 200 μM of H_2O_2 for 24 hr were senescent based on β -galactosidase staining whereas pre-treatment of hMSCs with 50 μM EGCG for 6 hr reduced senescence to 50% [36]. Control of oxidative stress generated during biomaterial transplantation or cell transplantation is important, and several studies have confirmed that tissue regeneration improves when antioxidants are used [37,38]. For example, silk fibroin nanofibers incorporating the antioxidant fenugreek (1:1) had 67% DPPH radical scavenging effect after 24 hr and improved wound healing in a rat full thickness excision model as compared to control and silk fibroin only [39]. In addition, anti-oxidative polyorganophosphazene (PATGP) microspheres fabricated using aniline tetramers and glycine ethyl ester co-substitution showed more than twice the DPPH, $\cdot\text{OH}$ scavenging activity and bone regeneration activity in a rat calvarial defect model after 8 and 16 weeks compared to PLGA and PAGP microspheres [40]. We found that the spread area of hADSCs cultured on Na-E and Mg-E surfaces was significantly greater than that on PCL film. H_2O_2 can form highly reactive hydroxyl radicals either spontaneously or through a Fenton reaction that causes oxidation, which directly attacks the cell walls (lipids) and DNA of hADSCs [41]. Therefore, it can be interpreted that the reduced spread area of damaged hADSCs may be due to the contraction of apoptotic cells, whereas cell adhesion and spread area was improved on EGCG-coated PCL film due to protective effect by ROS scavenging properties of EGCG. Phenols can react with the free radical species to delocalize unpaired electrons within the aromatic ring, giving phenol radicals that undergo resonance-stabilization within the molecule and thus, form stable intermediates [42]. H_2O_2 is a non-radical oxidizing agent that can decompose in physiological conditions to form highly reactive hydroxyl radicals ($\cdot\text{OH}$) [43,44]. The hydroxyl radicals cause cytotoxicity, and the resonance-stabilization by the aromatic ring of phenol can remove this hydroxyl radicals by aforementioned resonance-stabilization [42]. Given that, the delocalization of unpaired electrons within the phenolic structure of MPN coated on the surface may have removed cytotoxic hydroxyl radicals and produced stable H_2O and O_2 , improving cell viability.

3.3. Anti-apoptotic and anti-oxidative activities of EGCG-coated PCL film against H_2O_2

H_2O_2 treatment did not change the number of cells attached to the surfaces of Na-E and Mg-E, while it significantly decreased the number of cells attached to bare PCL film (Fig. 5b). In addition, the percentage of TUNEL-positive nuclei (%) was significantly greater for cells attached to PCL ($75 \pm 7\%$) than Na-E ($35 \pm 12\%$) and Mg-E ($10 \pm 10\%$) (Fig. 5a and c). H_2O_2 causes apoptotic DNA fragmentation, and EGCG has been shown to reduce the damage caused by H_2O_2 . For example, treatment of pancreatic alpha TC1-6

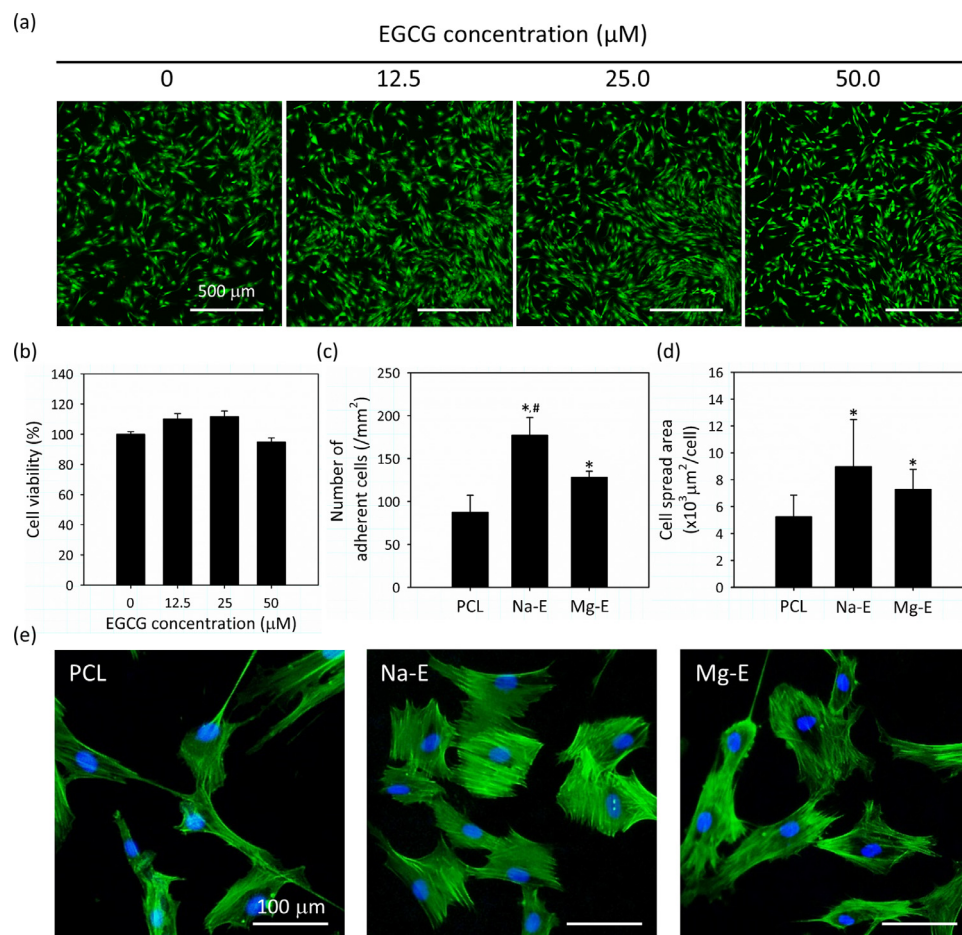


Fig. 3. EGCG cytotoxicity and attachment of hADSCs to PCL film. (a) LIVE/DEAD staining of hADSCs after 24 hr of EGCG treatment (0, 12.5, 25, 50 μM) and (b) MTT assay results. (c) The number of hADSCs attached to PCL, Na-E, and Mg-E after 1 day of culture. After F-actin staining, (d) cell spread area and (e) fluorescence were quantified (*, # $p < 0.05$).

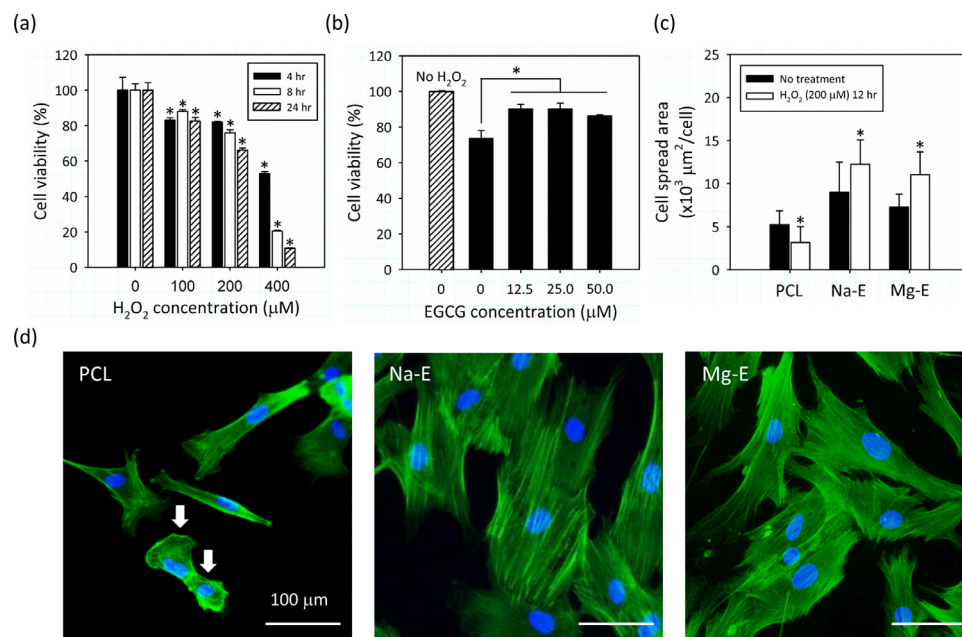


Fig. 4. Viability and morphological change of hADSCs after H_2O_2 treatment. (a) Viability of hADSCs after treatment with 0, 100, 200, or 400 μM H_2O_2 for 4, 8, and 24 hr. (b) Viability of hADSCs after 1 day of culture with 200 μM H_2O_2 + EGCG (0, 12.5, 25, 50 μM). After 12 hr of 200 μM H_2O_2 treatment of hADSCs cultured for 1 day on the PCL film surface, changes in (c) cell spread area and (d) fluorescence were quantified (*, # $p < 0.05$).

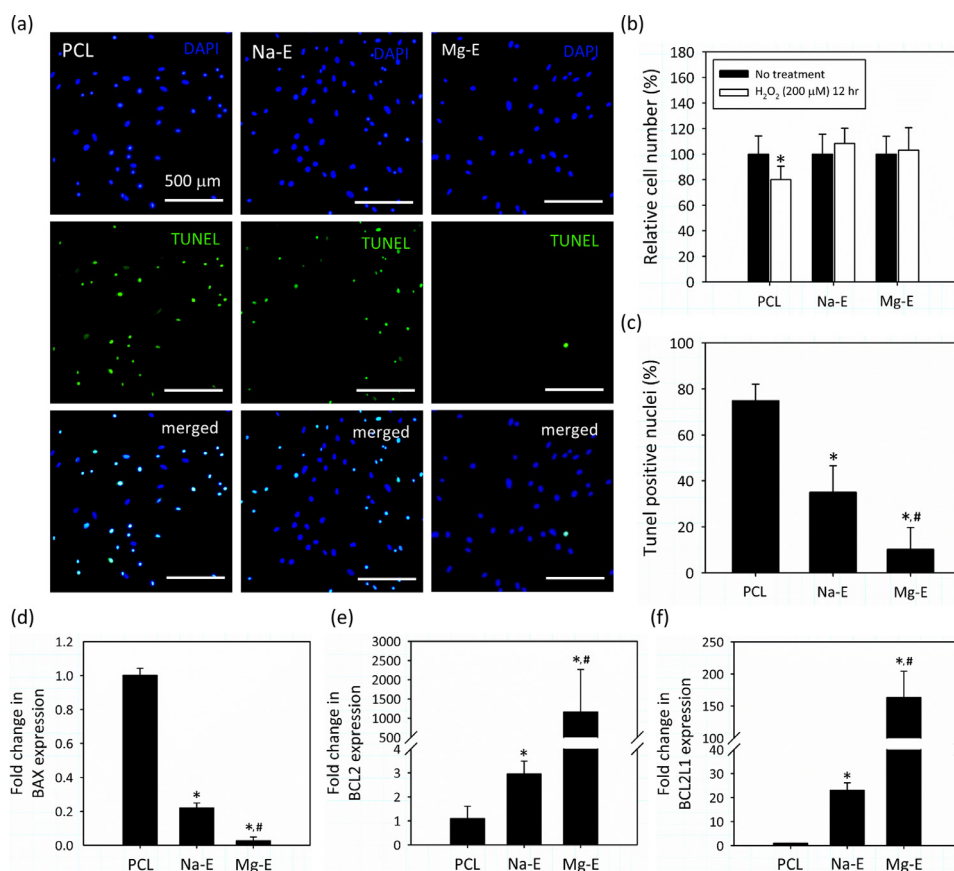


Fig. 5. EGCG-coated PCL film protects against H₂O₂-induced apoptosis. (a) Apoptotic signals of hADSCs cultured on the surface of PCL film after H₂O₂ treatment as measured by fluorescence microscopy after the TUNEL assay. (b) Change in relative cell number of hADSCs on PCL film after H₂O₂ treatment and (c) percentage of TUNEL-positive nuclei. Relative expression of the apoptosis-related genes (d) BAX, (e) BCL2, and (f) BCL2L1 in hADSCs (*, # $p < 0.05$).

cells for 4 hr with 100 μM H₂O₂ resulted in 90% TUNEL-positive nuclei, which was reduced to 70% and 5% after 1 h pre-treatment with EGCG 30 μM and 100 μM, respectively [45]. Our results suggest that EGCG coated on the polymer surface was also able to protect against H₂O₂-induced apoptosis in a dose-dependent manner. The expression of the apoptotic gene BAX in cultured hADSCs was significantly decreased in cells cultured on Na-E and Mg-E relative to bare PCL film, with the greatest decrease in expression of this gene observed for cells cultured on Mg-E (Fig. 5d). Expression of the anti-apoptotic genes BCL2 and BCL2L1 was significantly increased in Na-E and Mg-E with the highest expression observed for cells grown on Mg-E compared to bare PCL film (Fig. 5e and f). EGCG has previously been reported to have a modulatory effect on the expression of apoptosis-related genes; mouse vascular smooth muscle cells cultured in a conditioned environment with H₂O₂ showed significantly reduced expression of the pro-apoptotic genes caspase-3,8,9 and BAX and a significant increase in the expression of the anti-apoptotic gene BCL-2 after pre-treatment with EGCG [46]. Our results confirmed that the EGCG coating on the surface of PCL film was biologically active and influenced anti-oxidative signaling processes within hADSCs cultured on EGCG-coated film.

To investigate the H₂O₂ scavenging activity of the EGCG coating, PCL film and EGCG-coated PCL film were incubated in 100 μM H₂O₂ solution for 2 hr. H₂O₂ level was not reduced by the uncoated PCL film while levels of H₂O₂ were significantly decreased in solutions containing Na-E and Mg-E film to $97 \pm 1\%$ and $87 \pm 2\%$ of control levels, respectively (Fig. 6a). The relative MFI per cell in the DCFH-DA assay was significantly lower for cells cultured on Na-E and Mg-E film at $43 \pm 10\%$ and $24 \pm 5\%$ of the MIF per cell

observed for cells cultured on PCL film (Fig. 6b). Fluorescence microscopy revealed that hADSCs cultured on PCL film without H₂O₂ showed little fluorescence signal (Fig. 6c), while there was a significant increase in fluorescence signal after H₂O₂ treatment (Fig. 6d). In addition, fluorescence intensity was reduced in Na-E and Mg-E compared to PCL film (Fig. 6d). The expression of catalase, FOXO3, and GPX-1 was significantly increased in cells cultured on Na-E and highest for cells cultured on Mg-E compared to cells cultured on bare PCL film (Fig. 6e, f, and g). Polyphenols have been reported to eliminate H₂O₂ and related ROS. For example, real-time H₂O₂ monitoring of a skin membrane-covered oxygen electrode (SCOPE) revealed that 1 mM H₂O₂ was rapidly reduced after treatment with 0.1 mM polyphenols such as hydroquinone, quercetin, piceatannol, and resveratrol [47]. As described in Section 3.2, EGCG is expected to be able to remove H₂O₂ by resonance-stabilization mechanism, and our results suggest that EGCG retained its phenolic structure after coating. Furthermore, the EGCG coating appeared not only to remove ROS through direct anti-oxidative processes and resonance-stabilization, but also to regulate anti-oxidative enzyme expression in proportion to the TPC deposited on the film.

3.4. Characterization of EGCG-coated PLLA fibers

We next investigated the anti-oxidative protective effects of an EGCG coating on 3D spheroidal cultures of stem cells by preparing EGCG-coated PLLA fibers. According to the EGCG coating, the white color of PF turned brown in E-PF (Supplementary Fig. S5). SEM photographs (Fig. 7a) demonstrated that the smooth surfaces of PF became rough after coating with 48 ± 2 μg/mg EGCG, indicating

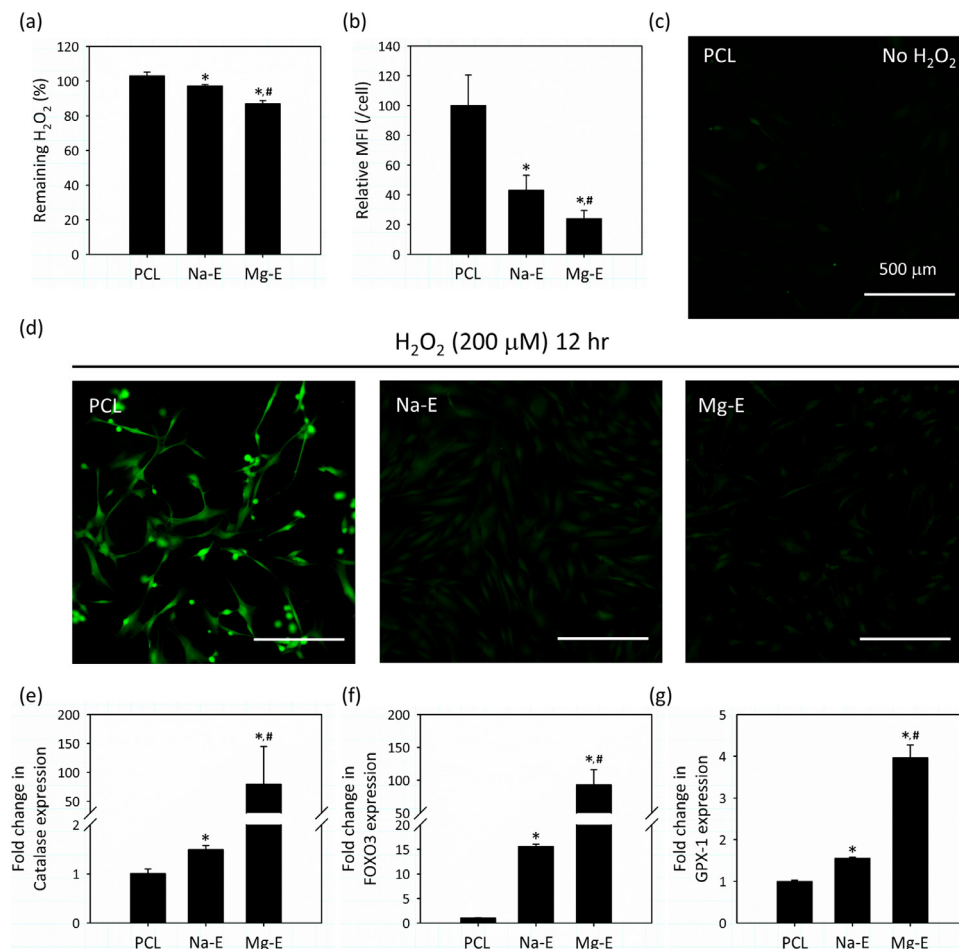


Fig. 6. Anti-oxidative activity of EGCG-coated PCL film in response to H₂O₂ treatment. (a) The amount of H₂O₂ remaining in D.W. after a 2 hr incubation with PCL film. (b) Intracellular ROS level of hADSCs cultured on PCL, Na-E, and Mg-E as determined by DCFH-DA and fluorescence images (c) without H₂O₂ treatment (d) with H₂O₂ treatment. Relative expression of anti-oxidative genes (e) catalase, (f) FOXO3, and (g) GPX-1 in hADSCs (*, # p < 0.05).

homogeneous deposition of EGCG (Fig. 7b). Ferric reducing activity [conversion rate (%)] increased up to $28 \pm 5\%$ as the percentage of fibers increased (Fig. 7c). Similarly, E-PF showed ABTS radical scavenging activity that increased in proportion to the amount of fibers, as shown in Fig. 7d. In an aqueous environment containing H₂O₂, E-PF effectively reduced H₂O₂ and thus decreased the amount of H₂O₂ significantly ($85 \pm 2\%$) as a function of fiber concentration (Fig. 7e). EGCG-coated fibers had no detrimental effect on cell viability (Fig. 7f). Intracellular ROS level (relative MFI) increased rapidly in the positive control (143 ± 17) treated with H₂O₂ compared to the negative control (100 ± 1) without H₂O₂ treatment. In the PF-treated group, fluorescence intensity per individual cell was not statistically different than that in the positive control (140 ± 8), but was significantly decreased in the E-PF treated group (110 ± 4) (Fig. 7g and h).

The aforementioned results suggest that EGCG directly removes ROS through phenolic π -electron and resonance-stabilization of catechin and gallol groups, and/or stimulates anti-apoptotic and anti-oxidative cell signaling pathways. Similarly, EGCG-coated fibers exhibited anti-oxidative properties as confirmed by Fe conversion, ABTS radical scavenging, and elimination of H₂O₂, indicating maintenance of the biological activity of EGCG independent of the surface chemistry of the underlying materials. In addition, E-PF effectively reduced the intracellular ROS level of hADSCs cultured on tissue culture plates. It has been reported that the level of intracellular ROS is indicative of cellular damage. For example,

the greater the intracellular ROS level of primary cardiac stem cells and embryonic stem cells, the more DNA damage was found [48]. The antioxidant curcumin (0–4 μ M) concentration-dependently decreased the DCF fluorescence intensity of L-6 myoblasts [49], while the antioxidant edaravone decreased the intracellular ROS level of human umbilical cord mesenchymal stem cells [11], indicating that these antioxidants can decrease the level of intracellular ROS. However, directly delivered soluble antioxidants were rapidly eliminated (8–90% removed within 24 hr) and showed decreased anti-oxidative activity *in vivo* [12]. Our results indicate coating the surfaces of biomaterials with EGCG can effectively remove ROS and potentially protect tissues against oxidative damage. It is known that soluble EGCG exhibits anti-oxidative properties in proportion to the concentration, but its use has been very limited in tissue engineering applications due to potential cytotoxicity at concentrations above 50 μ M [33,50]. When EGCG was coated on biomaterial, EGCG can be stably coated on the surface via MPN formation and could be maintained for a long time without cytotoxicity.

3.5. Spheroid formation with EGCG-fibers and their ability to protect hADSCs within the spheroids against H₂O₂

We next fabricated PLLA fibers for incorporation in stem cells spheroids and investigated their effects on 3D cultured stem cells. Similar to the aforementioned experiments, spheroids were exposed to 400 μ M H₂O₂ for 12 hr. Structural deformation was ob-

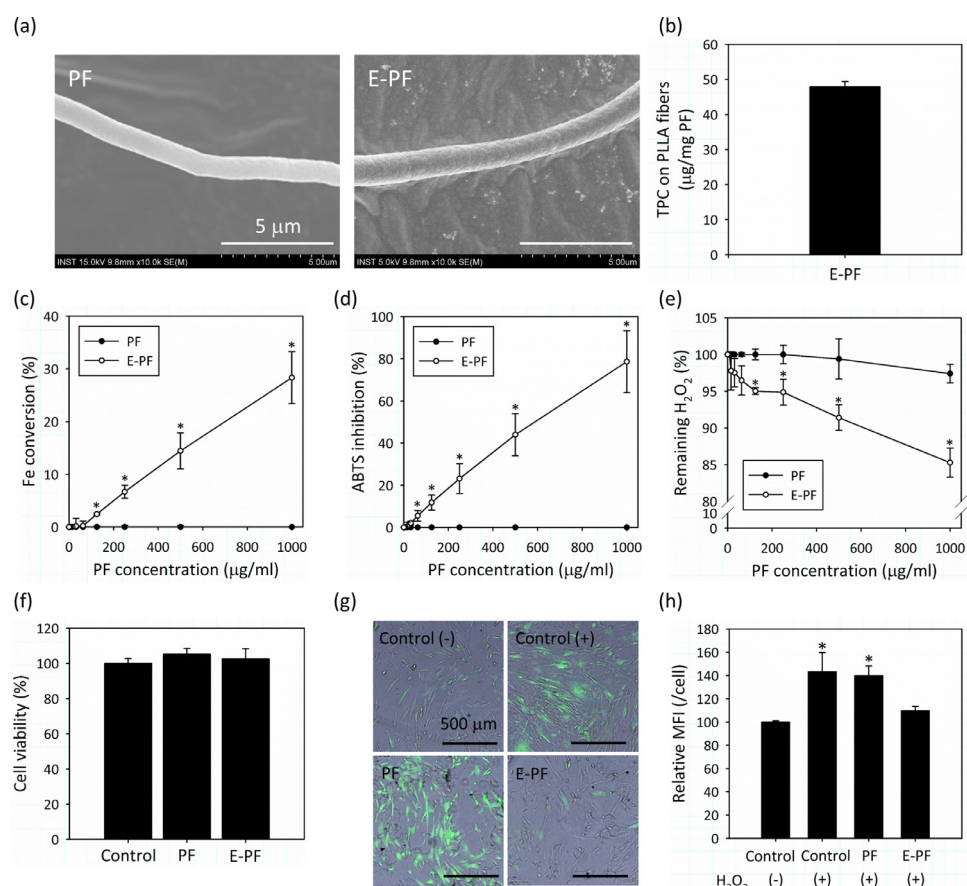


Fig. 7. Characterization of PLLA fibers (PF) and EGCG-coated PLLA fibers (E-PF). (a) SEM image of PF and E-PF surfaces. (b) The TPC on E-PF. The anti-oxidative activity of PF and E-PF was measured by (c) ferric reducing antioxidant power assay, and (d) ABTS radical scavenging assay. (e) The amount of H_2O_2 remaining after incubation of PF and E-PF in 100 μM of H_2O_2 in D.W. for 2 hr. (f) Viability of hADSCs cultured with PF or E-PF for one day. (g) Fluorescence images of intracellular ROS in hADSCs cultured with PF or E-PF in the presence of H_2O_2 for 1 day and (h) relative MFI per individual cell based on the DCFH-DA assay (* $p < 0.05$).

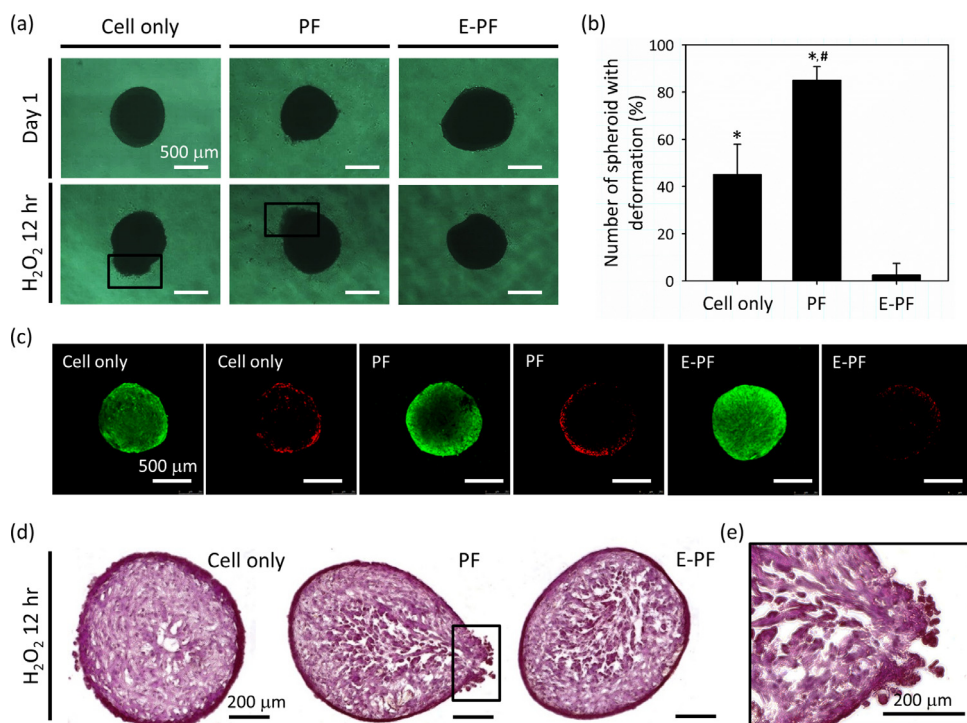


Fig. 8. Structural deformation of hADSCs spheroid in response to H_2O_2 exposure. (a) Phase contrast image of cell-only, PF, and E-PF spheroids and (b) percentage of deformed spheroids (%). (c) Survival of hADSCs in spheroids after LIVE/DEAD staining. (d) Internal morphology of spheroids as determined by H&E staining and (e) high resolution imaging of PF-incorporating spheroids (*, # $p < 0.05$).

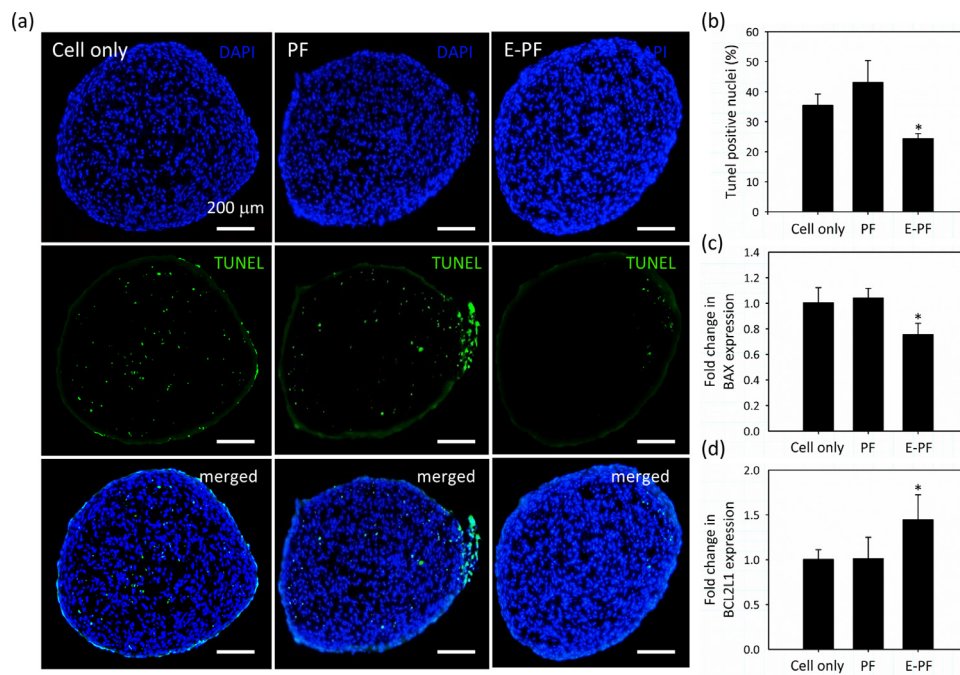


Fig. 9. Anti-apoptotic activity of EGCG-coated PLLA fibers incorporated in hADSC spheroids in response to H_2O_2 exposure. (a) Apoptotic signals of cell-only, PF, and E-PF spheroids and (b) percentage of TUNEL-positive nuclei (%). (c) Relative expression of (d) BAX and (f) BCL2L1 in cell-only, PF and E-PF spheroids (* $p < 0.05$).

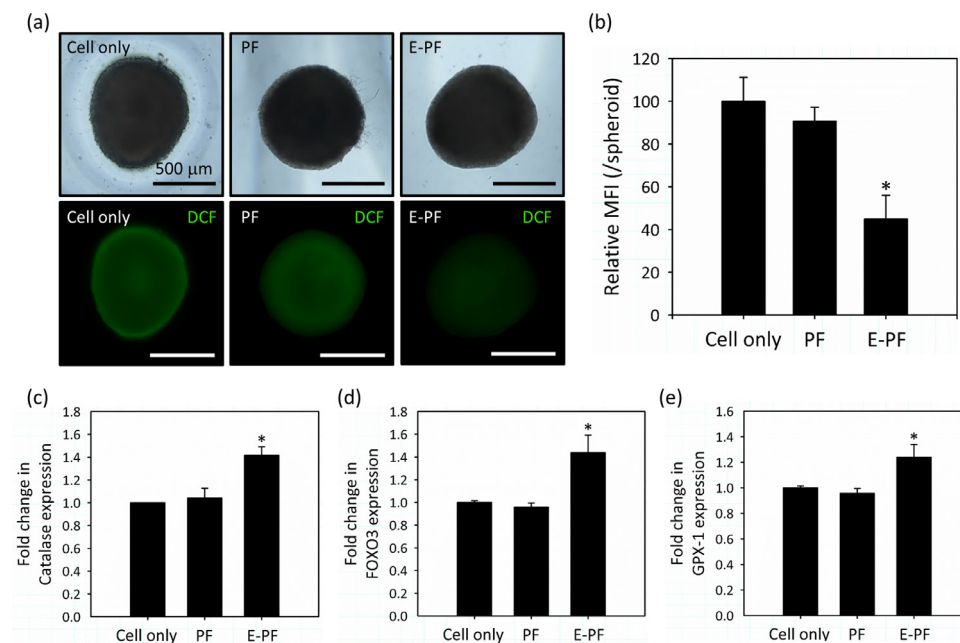


Fig. 10. Anti-oxidative activity of EGCG-coated PLLA fibers incorporated in hADSCs spheroids against H_2O_2 . (a) Intracellular ROS in spheroids were observed by fluorescence microscopy and (b) levels were quantified. Relative expression of anti-oxidative genes (e) catalase, (f) FOXO3, and (g) GPX-1 in cell-only, PF and E-PF spheroids (* $p < 0.05$).

served in the cell-only ($45 \pm 13\%$) and PF ($85 \pm 6\%$) spheroids, while there was no deformation of spheroids incorporating E-PF ($3 \pm 5\%$) (Fig. 8a and b). There were more dead cells in cell-only and PF spheroids than in E-PF spheroids (Fig. 8c). Histological analysis of cross-sectioned spheroids revealed that cells were homogeneously distributed and tightly integrated throughout the spheroids (Fig. 8d) while in the PF group, deformation of spheroids at the periphery was observed (Fig. 8e). Our results indicate that cell damage due to diffusion of H_2O_2 in the media was localized at the periphery of the PF spheroids. Deformation (%) was larger in the PF group than the cell-only group because the diffusion

of H_2O_2 was mitigated by the relatively loosely formed networks between cells and fibers. In fact, histological images revealed a thicker cellular layer at the periphery of cell-only spheroids while the outer cell layer became thinner in spheroids containing fibers (Fig. 8d). The protective function of E-PF against H_2O_2 was evident. Necrotic core formation or external damage can lead to contraction and eruption of spheroids and thus structural deformation of a spheroid can serve as an indicator of the integrity of the spheroid. For example, extensive DNA damage was observed in multicellular tumor spheroids (MCTSs) cultured with H_2O_2 (0.1–5 mM) for 1 hr, resulting in structural changes [51], while laser irradiation of MSC

spheroids caused destruction from the outside of the spheroid [52]. Our results indicate that the presence of E-PF protected against ROS damage.

There was a large number of TUNEL-positive nuclei in the PF group in the region of deformation (Fig. 9a), and the number of TUNEL-positive nuclei was significantly decreased in E-PF spheroids ($24 \pm 2\%$) compared to cell-only spheroids ($35 \pm 4\%$) and PF spheroids ($43 \pm 7\%$) (Fig. 9b). Expression level of the apoptotic gene BAX was significantly lower in E-PF spheroids than in cell-only and PF spheroids (Fig. 9c), and conversely, the expression of the anti-apoptotic gene BCL2L1 was significantly higher in E-PF spheroids than in cell-only and PF spheroids (Fig. 9d). Intracellular ROS levels in spheroids were determined after H_2O_2 treatment. The relative MFI per spheroid was 45 ± 11 for E-PF spheroids, which was significantly lower than that in PF spheroids (90 ± 7) and cell-only spheroids (100 ± 11) (Fig. 10a and b). Catalase, FOXO-3, and GPX-1 expression was significantly higher in E-PF spheroids than cell-only and PF spheroids (Fig. 10c, d, and e). Consistent with a previous report that intraperitoneal injection of EGCG (70 mg/kg) improved expression of the anti-oxidative enzyme GPX-1 in hepatic tissue [36], EGCG-coated fibers appear to modulate hADSC signaling in spheroids, resulting in improved anti-oxidative enzyme expression. Our research group has been investigating strategies to deliver instructive signals to cells inside spheroids by incorporating biofunctionalized fibers into the spheroids during spheroid generation. This approach could efficiently direct and modulate cellular activities within spheroids. For example, fibers have been functionalized with PDGF to improve the proliferation of hADSCs within spheroids [53] and to enhance osteogenic differentiation through adenosine immobilization and mineralization [54,55]. Our results suggest that incorporating EGCG-coated fibers in spheroids confers the spheroids with sustained anti-oxidative properties and modulates signaling by cells inside the spheroid.

4. Conclusions

In this study, PCL surfaces were coated with EGCG in the presence of Na^+ or Mg^{2+} cations via MPN formation, and the TPC was greater in the presence of Mg^{2+} than Na^+ . EGCG coating increased surface hydrophilicity and roughness proportional to the TPC. EGCG coating maintained its anti-oxidative properties as confirmed by Fe conversion and ABTS radical scavenging activity of hADSCs cultured on EGCG-coated surfaces in the presence of H_2O_2 . EGCG coating directly removed surrounding H_2O_2 and modulated signaling in hADSCs, leading to a decrease in expression of the apoptotic gene BAX and an increase in the expression of the anti-apoptotic genes BCL2 and BCL2-L1 as well as the anti-oxidative enzymes catalase, FOXO3, and GPX-1. Furthermore, EGCG-coated PLLA fibers exhibited anti-oxidative properties as evidenced by Fe conversion, ABTS radical scavenging activity, and removal of H_2O_2 . EGCG-coated PLLA fibers incorporated in stem cell spheroids reduced apoptotic signals in the presence of H_2O_2 -induced oxidative stress and induced anti-oxidative enzyme expression relative to cell-only spheroids and spheroids incorporating fibers without an EGCG coating. In conclusion, EGCG coating can effectively protect against ROS-induced oxidative damage, suggesting its potential versatility in various tissue engineering applications.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Acknowledgements

This research was supported by National Research Foundation of Korea (NRF) grants funded by the Korean government (MEST) (NRF-2019R1A2C2084965, NRF-2020R1A4A3078645).

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.actbio.2021.02.005.

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